miR-1236-3p represses the cell migration and invasion abilities by targeting ZEB1 in high-grade serous ovarian carcinoma

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Abstract. Ovarian cancer, particularly high-grade serous ovarian carcinoma (HG-SOC), is still the main cause of death among gynecological malignancies. However, the molecular mechanisms related to its malignant biological behavior are still unclear. Recent studies indicate that microRNAs (miRNAs) play an important role in tumor metastasis. Here, we report that miR-1236-3p expression was downregulated in HG-SOC when compared to that in normal fallopian tube tissue. Manipulation of miR-1236-3p significantly influenced the morphology, migration and invasion of ovarian cancer cell lines (A2780 and SKOV3). With dual-luciferase reporter assay, we demonstrated that miR-1236-3p binds to the 3’UTR of zinc-finger E-box binding homeobox 1 (ZEB1) mRNA, and functions as a negative regulator of ZEB1. Furthermore, we revealed that manipulation of miR-1236-3p modulates ZEB1 expression and influences expression of its downstream genes E-cadherin and N-cadherin at both the mRNA and protein levels. We also found an inverse relationship between miR-1236-3p and ZEB1 expression in the HG-SOC tissue samples. Taken together, our results indicate that miR-1236-3p regulates ovarian cancer metastasis by directly targeting ZEB1, and it may play an important role in the diagnosis and treatment of ovarian cancer.

Introduction

Even after decades of research investments, ovarian cancer remains the most lethal gynecological malignancy. The 5-year survival rate is only 44% in the USA (1). Due to histologic and molecular heterogeneity, epithelial ovarian cancer (EOC) is not usually considered a single entity. EOC can be divided into four major subtypes: serous, mucinous, endometrioid and clear cell adenocarcinomas. Serous adenocarcinoma is the most common among the subtypes and consists of low-grade and high-grade carcinomas. Most ovarian cancer deaths are associated with high-grade serous ovarian carcinoma (HG-SOC) (2), which originates from the fallopian tube fimbriae according to recent studies (3-7). Metastasis is the major cause for the high mortality rate. Therefore, it is crucial to clarify the molecular mechanisms that influence the metastasis of HG-SOC.

Epithelial-mesenchymal transition (EMT) is known as a key regulatory mechanism of migration and invasion in many types of cancers including ovarian cancer (8,9). EMT is a morphological change where cells switch from a polarized epithelial phenotype to a highly motile mesenchymal phenotype (10). During the process of EMT, cells lose epithelial adhesion molecules (such as E-cadherin) and acquire mesenchymal markers (such as N-cadherin), with increased migration and invasion. Several transcriptional factors such as Twist, ZEB and Snail have been reported as inducers of EMT (11). Among these factors, the zinc-finger E-box binding homeobox 1 (ZEB1) is known to be overexpressed in ovarian cancer and may directly repress the epithelial marker E-cadherin to induce EMT (12).

MicroRNAs (miRNAs) are a class of small (~22 nucleotides) non-coding RNAs that negatively modulate gene expression in a sequence-specific manner, and they are conserved in evolution (13). miRNAs play important roles in numerous biological processes, such as cell cycle, differentiation, proliferation, apoptosis and angiogenesis (14-16). The relationship between miRNAs and cancer was first discovered in chronic lymphocytic leukemia (17). Since then, numerous miRNAs have been found abnormally expressed in many types of cancers including ovarian cancer (18). EMT was also found to be modified by many miRNAs, such as the miR-200 family, miR-23b, miR-29b and miR-150 (19-23). In the present study, we examined the expression of miR-1236-3p in HG-SOC and normal fallopian tube tissues. We found that miR-1236-3p was downregulated in HG-SOC. We then investigated the effect of miR-1236-3p on tumor metastasis and showed that miR-1236-3p overexpression suppressed the migration and...
invasion of ovarian cancer cells by targeting EMT-inducer ZEB1.

Materials and methods

Patients and tissue samples. The present study consisted of 20 samples of HG-SOC and 12 normal fallopian tube (FT) tissues. All samples were collected between March and June 2013 at Qilu Hospital during surgery and were immediately stored at -80˚C. For the use of these samples, signed informed consent was obtained. All the diagnoses were confirmed by at least two pathologists.

Cell lines and culture conditions. Human ovarian cancer cell lines A2780 and SKOV3 were originally obtained from the American Type Culture Collection (ATCC, Rockville, MD, USA). Roswell Park Memorial Institute (RPMI)-1640 medium was purchased from Gibco-BRL (Rockville, MD, USA). Fetal bovine serum (FBS) was supplied by Haoyang Biological Manufacture Co. Ltd. (Tianjin, China). The human ovarian cancer cell lines A2780 and SKOV3 were cultured in RPMI-1640 medium supplemented with 10% FBS and 100 U penicillin-streptomycin at 37˚C in a humidified atmosphere containing 5% CO₂.

Synthesis and transfection of miRNA mimics and miRNA inhibitors. miRNA mimics and miRNA inhibitors (2’-O-methyl modified) were designed for in vitro transfection and its negative control (miR-nc) were designed and synthesized by RiboBio (Guangzhou, China). A2780 (3.5x10⁵) and SKOV3 (3.0x10⁵) cells were seeded into 6-well plates and incubated overnight. Transfection was performed using Lipofectamine 2000 (Invitrogen, Carlsbad, CA, USA) according to the manufacturer's instructions. After 6 h, the medium was replaced with fresh medium. During transfection, the medium was antibiotic-free.

Dual-luciferase reporter assay. The 3’-untranslated region (3’UTR) of ZEB1 mRNA containing the putative miR-1236-3p binding site was cloned into the pGL3 vector according to the manufacturer's instructions. The forward primer (5’-CGAGGCT CGACGACAGACAGGAA-3’) and the reverse primer (5’-CCCTCAGTAGTACGACGGTTGGA-3’) were used to amplify the 3’UTR of ZEB1. The forward primer contained a SacI restriction site and the reverse primer contained an XhoI site. The binding site of miR-1236-3p in the ZEB1 3’UTR was mutated by using primers: F, 5’-CAGGGCCTTAAAGG AAGCTGATTAAT-3’ and R, 5’-AGCGCCTGTATTGTTGAC TCTTGTAGT-3’. SKOV3 cells were plated at 1.5x10⁵/well in a 96-well plate one day before transfection. Cells were co-transfected with 50 ng of wild-type or mutant ZEB1 3’UTR, and 5 pmol of miR-1236-3p mimics or negative control. After 48 h, luciferase activity was measured using the Dual-Luciferase Reporter assay system (Promega).

RNA extraction and quantitative real-time PCR. Total RNA was extracted from cells using TRIzol reagent (Invitrogen) according to the manufacturer’s protocol. Then cDNA was synthesized using the PrimeScript RT reagent kit (Takara) by using the Applied Biosystems StepOnePlus™ Real-Time PCR System according to the manufacturer’s protocol. Glyceraldehyde-3-phosphate dehydrogenase (GAPDH) was used as the endogenous control.

Western blot analysis. The cells were washed 3 times with PBS chilled to 4˚C and lysed on ice in RIPA buffer (Shenneng Bocai, Shanghai, China) containing protease inhibitors (1 mM). The density of the protein was measured using the BCA protein assay kit (Merck, Darmstadt, Germany). The same amount of protein was separated by 5-10% SDS-PAGE and then transferred to a PVDF membrane (Millipore) using a semi-dry blotting apparatus (Bio-Rad Laboratories, Hercules, CA, USA). Membranes were blocked with 5% (w/v) non-fat milk in Tris-buffered saline with Tween-20 (TBST) (100 mM NaCl, 50 mM Tris and 0.1% Tween-20) at room temperature for 1 h. After the blocking step, the membranes were incubated overnight with primary antibodies (Cell Signaling Technology, Beverly, MA, USA) at 4˚C. The membranes were then washed 3 times with TBST and incubated with horseradish peroxidase-labeled secondary antibodies for 2 h. Signals were detected with an ECL system (Merck) according to the manufacturer's instructions. GAPDH was used as the loading control.

Invasion and migration assays. Invasion assays were performed in a Transwell system (24-wells, 8-µm pore size) coated with 2 mg/ml of Matrigel (both from BD Biosciences, Bedford, MA, USA). First, the cells were transfected with miR-1236-3p. After 48 h, the transfected cells were digested with trypsin, and 1.0x10⁵ cells were suspended in serum-free RPMI-1640 and seeded into the upper chamber of each well. The lower chamber was filled with RPMI-1640 supplemented with 20% FBS. Then the cells were fixed with formaldehyde, permeabilized with 100% methanol and stained with 0.5% crystal violet. The number of cells that had attached to the lower surface of the membrane were counted in 6 random fields under a light microscope and analyzed statistically. The cell migration assays were performed in a similar manner, except that the upper chambers were Matrigel-free.

Statistical analysis. All data are expressed as means ± standard deviation (SD). Student's t-test (paired, 2-tail) was employed to analyze the significance of two groups. P-value <0.05 was considered to indicate a statistically significant result. All of the experiments were repeated at least 3 times.

Results

Expression of miR-1236-3p is downregulated in HG-SOC. To confirm the expression of miR-1236-3p in HG-SOC tissue samples, we employed qRT-PCR to quantify and compare the miR-1236-3p expression levels in the HG-SOC (n=20) and FT (n=12) tissue samples. As shown in Fig. 1A, the expression of miR-1236-3p was generally decreased in the HG-SOC when compared to the FT samples (P<0.001). This suggests that miR-1236-3p acts as a tumor suppressor in HG-SOC.

Manipulation of miR-1236-3p-induced morphological change in SKOV3 and A2780 cells. To explore the influence of miR-1236-3p on ovarian cancer cells, we transfected SKOV3
and A2780 cells with miR-1236-3p mimics or inhibitors. As shown in Fig. 1C, SKOV3 cells treated with miR-1236-3p mimics presented a cobblestone-like appearance, while cells treated with the negative control exhibited a spindle-like morphology. We then inhibited the expression of miR-1236-3p in A2780 cells. A2780 cells transfected with the miR-1236-3p inhibitors were narrower than cells transfected with the negative control (Fig. 1D). As known, the classic morphological change correlated with EMT is the conversion from a rounded, epithelial-like form to a spindle-shaped, mesenchymal form (8,9). These changes indicated that the manipulation of miR-1236-3p inhibited or stimulated the process of EMT.

Migratory and invasive abilities of ovarian cancer cells are regulated by miR-1236-3p. To further confirm the role of miR-1236-3p in ovarian cancer cells, we used a two-chamber assay to evaluate the migratory and invasive capacities of SKOV3 and A2780 cells treated with miR-1236-3p inhibitors. As shown in Fig. 2, increased miR-1236-3p significantly suppressed migration and invasion of the ovarian cancer cells. In contrast, decreased miR-1236-3p promoted these abilities. Our results showed that miR-1236-3p greatly influenced migration and invasion of both ovarian cancer cell lines.

ZEB1 is targeted by miR-1236-3p. Based on the above findings, we hypothesized that miR-1236-3p regulates genes associated with EMT. Using online miRNA target prediction tools, such as microRNA.org, we found that the 3'UTR of ZEB1 mRNA contained a putative binding site for miR-1236-3p. To investigate the ability of miR-1236-3p to bind and regulate the 3'UTR of ZEB1, we performed the luciferase reporter assay. Wild-type or the mutant ZEB1 3'UTR sequence was cloned into the pGL3 vector (Fig. 3A). SKOV3 cells were co-transfected with miR-1236-3p (mimics or negative control) and the pGL3 vector (wild-type or mutant). After 48 h, cells were lysed to measure the luciferase activity using the Dual-Luciferase Reporter assay system. Overexpression of miR-1236-3p significantly reduced the luciferase activity in the wild-type (P<0.001) but not the mutant ZEB1 3'UTR (Fig. 3B). This demonstrated that ZEB1 was directly targeted by miR-1236-3p.

miR-1236-3p regulates the expression of ZEB1 and EMT-related genes. We then tested whether miR-1236-3p modulates the expression of ZEB1 and EMT-related genes in ovarian cancer. First, the SKOV3 and A2780 cell lines were transfected with miR-1236-3p mimics or miR-nc. The transfected cells were analyzed by qRT-PCR and western blotting. As shown in Fig. 4A and B, ZEB1 was downregulated by miR-1236-3p mimics at both the protein and mRNA levels. We also detected the expression of EMT-related markers E-cadherin and N-cadherin. The results showed that the expression of E-cadherin was upregulated and the N-cadherin expression was downregulated. Second, we downregulated the expression of miR-1236-3p by using miR-1236-3p inhibitors. As expected, the expression of ZEB1 and EMT-related genes showed an opposite pattern at the protein level (Fig. 4C) and mRNA level (Fig. 4D) compared to the previous assay. As known, a switch from epithelial marker E-cadherin to mesenchymal marker N-cadherin is a classical molecular change during EMT (8,9). Thus, our results indicated that miR-1236-3p may regulate the process of EMT.

Expression of ZEB1 protein is decreased in HG-SOC. Finally, we tested whether miR-1236-3p-induced ZEB1 suppression confirmed in our research was clinically relevant. As shown in Fig. 1B, we found that the expression of ZEB1 protein was
Figure 2. miR-1236-3p influences ovarian cancer cell migration and invasion. (A and B) A2780 and SKOV3 cells transfected with miR-1236-3p mimics (miR-1236-3p) showed reduced migration and invasion. (C and D) A2780 and SKOV3 cells transfected with miR-1236-3p inhibitors (anti-miR-1236-3p) showed enhanced migration and invasion. (E-H) The number of cells that migrated or invadod to the lower surface of the membrane were calculated. Data are presented as means ± SD. *P<0.05, **P<0.01.

Figure 3. ZEB1 is negatively regulated by miR-1236-3p. (A) Predicted miR-1236-3p binding site in the 3’UTR of ZEB1 and mutant ZEB1 3’UTR. (B) Dual-luciferase reporter assay showed that miR-1236-3p reduced the luciferase activity in wild-type 3’UTR of ZEB1 (WT-3’UTR) but not in the mutant ZEB1 3’UTR (mut-3’UTR). Data are expressed as the means ± SD. ***P<0.001. ZEB1, zinc-finger E-box binding homeobox 1; 3’UTR, 3’-untranslated region.
generally upregulated in HG-SOC (n=8) when compared to that in the FT (n=8) samples, and its expression was inversely correlated with miR-1236-3p. Taken together, our findings revealed that miR-1236-3p downregulation induced overexpression of ZEB1, and consequently influenced migration and invasion of HG-SOC cells.

Discussion

Early-stage ovarian cancer has few visible symptoms; therefore, most patients are diagnosed at advanced stages of disease when cancer cells have already spread (24). Current treatments (surgery, radiation and chemotherapy) are relatively ineffective for advanced ovarian cancer. Most of these patients will eventually relapse at metastatic sites. Thus, it is vital to understand the molecular mechanisms of metastasis. The present study showed that miR-1236-3p expression was decreased in HG-SOC tissue samples. Functional studies demonstrated that decreased miR-1236-3p enhanced ovarian cancer cell migration and invasion in vitro. These findings suggest that miR-1236-3p may play a potential inhibitory role in HG-SOC, and loss of miR-1236-3p may be critical for HG-SOC metastasis.

According to previous studies, miRNAs are highly tissue specific and function as tumor suppressors or oncogenes (25). The diagnostic, therapeutic and prognostic potential of miRNAs in cancer is promising (26). The miR-200 family plays an important role in the regulation of EMT by targeting the mRNA of ZEB1 and ZEB2. E-cadherin expression was also found to be correlated with miR-200 family expression in tissue samples from ovarian cancer patients (19,20). miR-1236-3p is reported to be involved in the regulation of VEGFR-3 and TLR4 (27,28). Meanwhile, TLR4 and VEGFR-3 are both associated with metastasis and the poor survival of ovarian cancer patients (29-32). Therefore miR-1236-3p may be relevant to the prognosis of ovarian cancer.

EMT facilitates the ability of cancer cells to detach themselves from primary lesions, migrate to distant organs or to invade adjacent tissue, and eventually form tumor metastases. EMT is considered to be the most important step in the progression of cancer from primary tumors to other organs. Thus, blocking of EMT is an efficient approach to inhibit the spread of cancer. ZEB1 has been previously reported to be involved in cancer progression, and is known as an important transcriptional repressors of E-cadherin. In the present study, we found that miR-1236-3p may influence the process of EMT in vitro. We demonstrated that overexpression of miR-1236-3p suppressed ZEB1 expression and the motility of ovarian cancer cells, while downregulation of miR-1236-3p promoted them. Based on our findings, further studies may be required to verify the function of miR-1236-3p in vivo and whether the expression of miR-1236-3p is associated with the overall survival of HG-SOC patients.

In conclusion, the present study first demonstrated that miR-1236-3p directly targets the EMT-inducer ZEB1 and is downregulated in HG-SOC. Manipulation of miR-1236-3p regulated the invasion and migration of ovarian cancer cells. In addition, the present study indicates that miR-1236-3p may be a potential target for the prognosis and treatment of HG-SOC.

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