Upregulation of NEK2 is associated with drug resistance in ovarian cancer

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Abstract. NEK2 [NIMA (never in mitosis gene A)-related expressed kinase 2] is associated with various biological behaviors in the development of cancer, while research concerning its association with drug resistance is limited. The association of NEK2 with drug resistance in ovarian cancer has not yet been reported. In the present study, on the basis of microarray results from Oncomine and the GEO Profiles online database, we revealed that NEK2 mRNA expression in ovarian cancer tissues is upregulated. In addition, its expression in drug-resistant ovarian cancer cells was upregulated when compared with expression with their sensitive or parental counterparts. Finally, we performed a comprehensive bioinformatic analysis consisting of protein/gene-protein/gene interaction network, annotation of biological processes and microRNA-mRNA interaction analysis. We observed that NEK2 directly or indirectly interacts with a number of genes, proteins, microRNAs and biological processes associated with drug resistance in ovarian and other types of cancer. These results indicate that NEK2 contributes to drug resistance in ovarian cancer and it may be an important therapeutic target.

Introduction

Ovarian cancer is the leading cause of death among malignancies of the female reproductive system, with a high rate of mortality worldwide. Early-stage ovarian cancer is frequently asymptomatic and difficult to detect; thus, most patients are in advanced stages [International Federation of Gynecology and Obstetrics (FIGO) stage III and IV] at the time of initial diagnosis (1), and 5-year survival rates are less than 40% with only modestly improved survival noted over the past 40 years (2). The current therapy for ovarian cancer is debulking surgery followed by cisplatin-centered chemotherapy (3). Although cisplatin-centered chemotherapy achieves a complete response rate of 40 to 60% in advanced ovarian cancer patients (4), long-term survival remains poor as a result of recurrence and emergence of drug resistance that finally leads to fatal disease (5).

Drug resistance, both intrinsic and acquired, results from a variety of factors including individual variations in patients and somatic cell genetic differences in tumors (6). A number of factors such as decreased cell-associated drugs, altered drug inactivation, increased DNA damage tolerance/repair, increased anti-apoptotic regulator activity and growth factor receptor deregulation are considered to be responsible for drug resistance in ovarian cancer (7,8). However, regardless of the mechanisms, abnormal expression of drug resistance-related genes often plays an important role in drug resistance. And among all of these drug resistance-related genes, oncogenes are clearly the key players.

Oncogenes refer to genes whose activation can contribute to the development of cancer (9) and many are involved in drug resistance in varied types of cancers. For example, the overexpression and activation of the c-myc oncogene is associated with drug resistance in small cell lung carcinoma (10); the LRF oncogene is a survival factor in chondrosarcoma and contributes to tumor malignancy and drug resistance (11) and the STAT3 oncogene is a predictive marker of drug resistance in cancer (12). In ovarian cancer, a total of 13 oncogenes including CCNE1, PIK3CA, RAB25, MYC, PRKCI, FGFI, NOTCH3, PIK3R1, AKT2, EGFR, ERBB2, KRAS and AURKA are associated with cancer development (13), and several genes such as KRAS (14), ERBB2 (15), PIK3CA (16,17) and PIK3R1 (18) are involved in drug resistance.

NEK2 [NIMA (never in mitosis gene A)-related expressed kinase 2], a serine/threonine centrosomal kinase, is highly expressed and activated during the S and G2 phases of the cell cycle (19). NEK2 has emerged as an important oncogene due to its regulatory role in mitosis (20). NEK2 has been proven to play pivotal roles in the development of several types of cancers, while its relationship with ovarian cancer is rare.

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More recently, NEK2 has become important since two studies indicate that high expression of NEK2 induces drug resistance in multiple myeloma (21,22). However, despite these studies, the research on NEK2 in regards to drug resistance in cancers is still limited, and its relationship with drug resistance in ovarian cancer has never been reported. In the present study, on the basis of a comprehensive bioinformatic analysis, for the first time, we report that NEK2 may contribute to drug resistance in ovarian cancer.

Materials and methods

The target gene, NEK2, which is closely associated with drug resistance in multiple myeloma (21,22) was selected for bioinformatic analysis.

Databases. The microarray data of NEK2 in ovarian cancer tissues was retrieved from the Oncomine online database (https://www.oncomine.org/resource/main.html); the microarray data of NEK2 in ovarian cancer cells was retrieved from the Gene Expression Omnibus (GEO) (http://www.ncbi.nlm.nih.gov/geo/). The protein/gene-protein/gene interaction network was generated using the GeneMANIA online tool (http://www.genemania.org/); annotation of biological processes was performed using the COREMINE online tool (http://www.coremine.com/medical/). The microRNAs (miRNAs) targeted to the gene were predicted by miRWalk online tool which included 7 prediction tools (DIANAmT, miRanda, miRD, miRWalk, RNAhybrid, PICTAR5, Targetscan) (http://www.umm.uni-heidelberg.de/apps/zmf/mirwalk/).

NEK2 expression in ovarian cancer cells retrieved from the GEO database was normalized. Unpaired, two-tailed t-test assuming homogeneity of the variances was performed with Excel software.

Results

Function of NEK2 in cancers. Genetic screening for cell division cycle mutants in the filamentous fungus Aspergillus nidulans resulted in the discovery of never in mitosis A (NIMA), a serine/threonine kinase that is essential for mitotic entry. Since then, NIMA-related kinases (NEKs) have been identified in most eukaryotes, including humans in which 11 genetically distinct proteins named NEK1 to NEK11 have been discovered (27). The NEKs play crucial roles in several aspects of mitotic progression, such as chromatin condensation, nuclear envelope breakdown, spindle assembly checkpoint signaling and cytokinesis. Of the human NEKs, NEK2 is the most closely related to Aspergillus NIMA and is the first NEK which has been relatively well studied (27). Similar to NIMA, NEK2 is a cell cycle regulated kinase, with a peak of expression and activity in the S and G2 phase of the cell cycle, and it is a core component of the human centrosome (28,29). NEK2 also contributes to the establishment of the microtubule-based mitotic spindle (27).

It has been proven that the NEK2 protein is elevated 2- to 5-fold in cell lines derived from a wide range of human tumors including those of cervical, ovarian, breast, prostate and leukemic origin (30), suggesting its potential roles in cancer development. The role of NEK2 in breast cancer has been extensively studied. In various human breast cancer cell lines, suppression of NEK2 was found to induce aneuploidy and cell cycle arrest resulting in cell death. Significantly, the breast cancer cell line which was most sensitive to NEK2 depletion was of the triple negative breast cancer subtype. These results indicate that NEK2 plays crucial roles in breast cancer growth at primary and secondary sites (31). Similarly, Wang et al (32) indicated that the abnormal expression of NEK2 and β-catenin may be one of the mechanisms for tumorigenesis, particularly for abnormal tumor proliferation, and the cytoplasmic expression of NEK2 is associated with both tumor grade and tumor size. These results suggest that NEK2 and β-catenin may represent new potential targets for therapeutic intervention. Moreover, a study indicated that the genetic polymorphisms of NEK2 are related to breast cancer susceptibility in Chinese Han women (33). In addition, the NEK2C, which is a splice variant of NEK2, which was found to have significantly upregulated expression in breast cancer cell lines as well as in human primary breast cancer tissue, plays a crucial role in breast cancer development and NEK2C inhibition may be a useful therapeutic target for human breast tumors (34). In addition to the important roles in breast cancer, NEK2 also plays a role in other types of cancer. For instance, NEK2 is shown to be involved in the development of lung cancer induced by smoking and affects patient survival (35). In cervical cancer, NEK2 was induced in 102 cancer biopsies when compared with 24 normal controls, indicating its potential roles in cervical cancer (36).

The role of NEK2 in ovarian cancer is less understood. Limited studies indicate that NEK2 is overexpressed in ovarian cancer cells (30) and its expression is regulated by NR2F2 which is associated with a significantly shorter disease-free interval (37), yet no further research has been reported. Likewise, the relationship between NEK2 and drug resistance is unclear, with only several studies indicating its drug resistance-related functions. The combined use of both NEK2 siRNA and cisplatin inhibited cell proliferation and induced apoptotic cell death in vitro, suggesting that NEK2 may be associated with drug resistance in colorectal cancer (38). Similarly, the combination of NEK2 siRNA with paclitaxel and doxorubicin was found to improve the sensitivity of breast cancer cells during chemotherapy treatment (20). More recently, two studies strongly support the idea that NEK2 contributes to drug resistance in cancers. Zhou et al (21) and Harrison (22) revealed that high expression of NEK2 induced drug resistance mainly through activation of efflux pumps and it may be a strong predictor of drug resistance and poor prognosis in multiple myeloma and other types of cancers.

Taken together, NEK2, an oncogene, plays crucial roles in the development of various types of cancers, but related studies in ovarian cancer are rare. In addition, although several studies have revealed the drug resistance-related functions of NEK2 in several cancers, studies in this area are limited and its association with drug resistance in ovarian cancer has never been reported.

Expression of NEK2 in ovarian cancer and drug-resistant cells. NEK2 is considered as an oncogene and is overexpressed in various tumors. Thus, as an oncogene, NEK2 should be
overexpressed in ovarian cancer when compared to the normal control and should be upregulated in drug-resistant cells when compared to sensitive cells. Based on microarray data retrieved from the Oncomine online database, we revealed that among 4 independent microarray analyses which covered a total of 759 ovarian cancer tissues and 26 normal controls, NEK2 was overexpressed 1.687- to 5.754-fold in ovarian cancer tissues when compared with the normal controls (Fig. 1A). Regarding two microarray data sets from the GEO Profiles database, NEK2 was found to be elevated 1.5- to 1.7-fold in cisplatin-resistant A2780 ovarian cancer cells when compared with the sensitive counterparts (Fig. 1B). These results suggest that NEK2 may be involved in the development of ovarian cancer, including the regulation of drug resistance.

**Functional prediction and analysis based on protein/gene-protein/gene interaction.** The protein/gene interaction of NEK2 with other proteins/genes was analyzed using GeneMANIA online tool. As shown in Fig. 2, NEK2 has direct interactions with 8 proteins/genes. Among these, NEK2 was co-expressed with RADS1, BRCA2, PTEN and DAPK1, was co-expressed and had genetic interaction with BRCA1, was co-expressed and shared protein domains with CDK2, shared protein domains with CHEK2 and had genetic interactions with NOM1. With the exception of NOM1, the other 7 proteins/genes are all associated with drug resistance in ovarian cancer.

BRCA1 and BRCA2 proteins are critically important for the repair of double-strand breaks (DSB) by homologous recombination (HR) (39). They cooperate in DNA damage responses in a PALB2-dependent manner and have important implications for the genesis of ovarian cancer and for chemotherapy with DNA interstrand cross-linking agents (40). The mRNA expression of BRCA1 has a negative correlation with the clinical sensitivity of platinum-based chemotherapy (41). In addition, BRCA1 is positively correlated with MDR1 which is significantly involved in drug resistance and disease progression (42). In regards to BRCA2, previous research indicates that the functional restoration of BRCA2 due to a secondary BRCA2 mutation is involved in acquired drug resistance of BRCA2-mutated ovarian carcinomas (43). These results implicate the important role of BRCA1 and BRCA2 in drug resistance in ovarian cancer. Rad51 functionally interacts with BRCA1 in the meiotic and mitotic cell cycles (44), and, on the basis of an interaction network (Fig. 2), Rad51 co-expressed, physically interacted and shared pathways with BRCA1, and co-expressed, and shared pathways with BRCA2. These results indicate that Rad51 which is closely associated with BRCA1 and BRCA2 may also have drug resistance-related functions in ovarian cancer. The PTEN tumor-suppressor is a central negative regulator of the PI3K/AKT signaling cascade and suppresses cell survival as well as cell proliferation. It is found to be either inactivated or mutated in various human malignancies. Previously studies suggest that PTEN may be involved in drug resistance via the PI3K/AKT pathway and the p53-mediated apoptotic cascade. Reduction in PTEN expression is found to result in the development of drug resistance in OVCAR-3 cells and the alterations conferred resistance to cisplatin through the activation of PI3K/Akt and the inhibition of Bax translocation (17). Further research indicates that overexpression of PTEN reverses chemoresistance to cisplatin in human ovarian cancer cells through inactivation of the
PI3K/AKT cell survival pathway and may serve as a potential molecular target for the treatment of chemoresistant ovarian cancer (45). Moreover, overexpression of PTEN upregulates p53 content and increases the sensitivity of chemoresistant cells to cisplatin-induced apoptosis without detectable changes in the levels of phosphorylated Akt, suggesting that PTEN may be involved in drug resistance through a p53-mediated apoptotic cascade independent of the PI3K/Akt pathway (46). CHEK2 is one of the critical kinases governing cell apoptosis, cell cycle checkpoint and DNA damage repair. In ovarian cancer cells, CHEK2 is degraded at the protein level in response to cisplatin through the ubiquitin-proteasome pathway, suggesting that degradation or decreased expression of CHEK2 is partially responsible for chemoresistance (47). The expression of DAPK1 and DAPK2 is altered in multi-drug-resistant gastric cancer cell lines and thus these genes may be an integral part of the mechanisms responsible for chemoresistance (48). In ovarian cancer, DAPK1 has direct interactions with FBXO32, PDCD4, PTEN, TP53 and TP73, and has indirect interactions with BRCA1, BRCA2, CDKN1A, IL24, MLH1 and SULF1, which are all involved in drug resistance in ovarian cancer, suggesting its potential role in drug resistance in ovarian cancer (49). As for CDK2, a previous study revealed that cisplatin consistently induces transient S-phase arrest by inhibiting the CDK2/cyclin A complex in the S-phase at 12 h after treatment with cisplatin or DAP in combination with the mitotic inhibitor nocodazole and also potently inhibits G1-phase CDK2/cyclin E activities at 18 h (50), indicating that CDK2 can directly respond to cisplatin in ovarian cancer cells.

In addition to those with direct interactions with NEK2, there were other proteins/genes in the network which indirectly interacted with NEK2 (Fig. 2). Among those, MLH1 (51,52), PDCD4 (53), TP53 (54), TP73 (55), WWOX (56), CDKN1A (57), PLAGL1, PYCARD, APC and TP63 (49), PALB2 (40), ABCC3 (58) and PMS2 (59) are associated with drug resistance in ovarian cancer.

Collectively, based on the protein/gene interaction network analysis, 8 proteins/genes were identified which directly interact with NEK2, and 7 of them were found to contribute to drug resistance in ovarian cancer; 24 proteins/genes were found to indirectly interact with NEK2 and 13 of them were found to be associated with drug resistance in ovarian cancer. These results indicate a potential role of NEK2 in drug resistance in ovarian cancer.

Functional annotation of NEK2 with its interacting proteins/genes was performed to further reveal the relationship of NEK2 with drug resistance. As shown in Table I, the main functions of NEK2 with its interacting proteins/genes are cell cycle-related and microtubule-related functions, which were both proven to be associated with drug resistance in ovarian and other cancer types. Cell cycle-mediated drug resistance is best described as a relative insensitivity to a chemotherapeutic agent because of the position of the cells in the cell cycle. The cell cycle is closely involved in the chemosensitivity to combination chemotherapy, and the chemotherapeutic agents correlated with cell cycle events include taxanes, platinum, camptothecin and fluorouracil (60). It has been proven that the cell cycle is closely associated with drug resistance in ovarian cancer. Integration of DNA methylation and gene expression reveals specific platinum resistance-related signaling pathways in ovarian cancer, which include cell growth-promoting pathways, PI3K/Akt and cell cycle progression (61). Moreover, numerous genes participate in drug resistance through regulation of the cell cycle. For example, Phb1 induces G0/G1 phase cell cycle arrest and promotes cancer cell survival, and silencing of Phb1 expression is a valuable therapeutic approach for chemoresistant ovarian cancer by increasing the sensitivity of cancer cells to apoptosis (62). Likewise, comprehensive bioinformatic analysis revealed that 15 tumor-suppressor genes (TSGs) associated with drug resistance in ovarian cancer perform their drug resistance-related functions through 5 pathways including...
In addition, the cell cycle is also involved in drug transportation in cancer. Multidrug resistance driven by ABC membrane transporters is one of the major reasons for treatment failure in human malignancies, and modulation of MDR by cell cycle-related factors has been observed in MCF-7 breast cancer cells (63).

Microtubules are intracellular tubular structures found in all eukaryotic cells. Microtubules have various functions including organization of intracellular structure, cell division and intracellular transport (64). However, abnormal changes in microtubules lead to drug resistance in cancers. For instance, RASSF1A, tumor-suppressor protein, forms an endogenous complex with tubulin and promotes the stabilization of microtubules. Previous research revealed a strong correlation between the reduced relative expression of RASSF1A and taxol resistance in primary ovarian cancer. The reason is that
the loss of RASSF1A expression sensitizes cells to microtu-
bume destabilizing stimuli finally resulting in the development
of drug resistance (65). In ovarian cancer treatment, anticancer
drugs including paclitaxel, epothilone B (EpoB) and discoder-
molide all specifically interfere with microtubules and arrest
cells in the G2/M phase of the cell cycle (66).

Collectively, on the basis of functional annotation, the two
ovarian cancer drug resistance-related functions, including
cell cycle- and microtubule-related functions, were anno-
tated to be the major functions of NEK2 with its interacting
proteins/genes of which most are associated with drug resis-
tance in ovarian cancer (Fig. 2). These results suggest that
NEK2 may contribute to drug resistance in a cell cycle- and/or
microtubule-mediated manner in ovarian cancer.

Functional prediction and analysis based on annotation of
biological processes. The biological process annotation was
performed using COREMINE online database/tool. As shown
in Fig. 3, a total of 14 biological processes were annotated
with NEK2, ovarian cancer and drug resistance (p<0.00265),
which suggested that, on the one hand, those biological
processes contribute to the development of ovarian cancer and
drug resistance, and on the other hand, NEK2 was associated
with these biological processes and it may be a regulator for
those processes. Taken together, the annotation of biological
processes suggested that NEK2 may contribute to drug
resistance via 14 biological processes in ovarian cancer. The
14 biological processes may be mainly divided into two groups
including growth/death-related processes which include cell
growth, cell division, cell proliferation, apoptosis and cell
death and cell cycle-related processes including cell cycle
arrest, regulation of cell cycle, mitosis, the cell cycle, S and
G2 phase. These results indicate that NEK2 may participate
in the regulation of ovarian cancer drug resistance mainly
through the cell cycle, cell growth and death, in particular,

through the cell cycle. This conclusion was consistent with the
result of our functional annotation (Table 1).

Functional prediction and analysis based on miRNA-mRNA
interaction. As shown in Table II, the top 10 miRNAs targeted
to NEK2 were predicted by 7 prediction tools, and the drug
resistance-related functions of these miRNAs were reviewed
and integrated. Among the top 10 miRNAs, 7 are associated
with drug resistance in ovarian and other cancers. For instance,
the expression levels of miR-27a were found to be increased in
paclitaxel-resistant ovarian cancer cell line A2780/taxol when
compared with its parental line A2780. Transfection of A2780/
taxol cells with inhibitors of miR-27a enhanced the sensitivity
of A2780/taxol cells to paclitaxel and increased paclitaxel-
induced apoptosis, which suggest that the deregulation of
miR-27a may be involved in the development of drug resistance
in ovarian cancer (67). miRNA expression profiles revealed
that miR-27b is downregulated in most drug-resistant Ehrlich
ascites tumor cells, suggesting this miRNA may be involved in
drug resistance (68), even though no further results have been
reported. miR-150 was found to have low expression in most
ovarian tumors with significantly increased expres-

Table II. The top 10 miRNAs targeted by NEK2 as predicted by miRNA-mRNA interactions and their drug resistance-related
functions in cancer.

<table>
<thead>
<tr>
<th>microRNA</th>
<th>microRNA-mRNA prediction tool</th>
<th>Drug resistance-related functions of the microRNAs in cancers (ref.)</th>
</tr>
</thead>
<tbody>
<tr>
<td>hsa-miR-27a</td>
<td>a, DIANAmT; b, miRanda; c, miRDB; d, miRWalk; e, RNAhybrid; f, PICTAR5; g, Targetscan</td>
<td>Drug resistance in ovarian cancer (67)</td>
</tr>
<tr>
<td>hsa-miR-27b</td>
<td>a, DIANAmT; b, miRanda; c, miRDB; d, miRWalk; e, RNAhybrid; f, PICTAR5; g, Targetscan</td>
<td>Drug resistance in Ehrlich ascites tumor cells (68)</td>
</tr>
<tr>
<td>hsa-miR-150</td>
<td>a, DIANAmT; b, miRanda; c, miRDB; d, miRWalk; e, RNAhybrid; f, PICTAR5; g, Targetscan</td>
<td>Drug resistance and metastasis in ovarian cancer (69)</td>
</tr>
<tr>
<td>hsa-miR-374a</td>
<td>a, DIANAmT; b, miRanda; c, miRDB; d, miRWalk; e, RNAhybrid; f, PICTAR5; g, Targetscan</td>
<td>Drug resistance in head and neck squamous cell carcinoma cells (70)</td>
</tr>
<tr>
<td>hsa-miR-374b</td>
<td>a, DIANAmT; b, miRanda; c, miRDB; d, miRWalk; e, RNAhybrid; f, PICTAR5; g, Targetscan</td>
<td>No reported functions in cancer</td>
</tr>
<tr>
<td>hsa-miR-630</td>
<td>a, DIANAmT; b, miRanda; c, miRDB; d, miRWalk; e, RNAhybrid; f, PICTAR5; g, Targetscan</td>
<td>Drug resistance in non-small cell lung cancer (71)</td>
</tr>
<tr>
<td>hsa-miR-24-2</td>
<td>a, DIANAmT; b, miRanda; c, miRDB; d, miRWalk; e, RNAhybrid; f, PICTAR5; g, Targetscan</td>
<td>Drug resistance in human cancer cells (MCF-7 and HeLa) (72)</td>
</tr>
<tr>
<td>hsa-miR-216b</td>
<td>a, DIANAmT; b, miRanda; c, miRDB; d, miRWalk; e, RNAhybrid; f, PICTAR5; g, Targetscan</td>
<td>Cell proliferation, invasion and tumor growth and cellular senescence in nasopharyngeal carcinoma and colorectal cancer (74,75)</td>
</tr>
<tr>
<td>hsa-miR-128</td>
<td>a, DIANAmT; b, miRanda; c, miRDB; d, miRWalk; e, RNAhybrid; f, PICTAR5; g, Targetscan</td>
<td>Drug resistance in breast cancer (73)</td>
</tr>
<tr>
<td>hsa-miR-486-5p</td>
<td>a, DIANAmT; b, miRanda; c, miRDB; d, miRWalk; e, RNAhybrid; f, PICTAR5; g, Targetscan</td>
<td>Cell migration and invasion in non-small cell lung cancer (76)</td>
</tr>
</tbody>
</table>

The miRNA-mRNA analysis tools: a, DIANAmT; b, miRanda; c, miRDB; d, miRWalk; e, RNAhybrid; f, PICTAR5; g, Targetscan; NEK2,
NIMA (never in mitosis gene A)-related expressed kinase 2.
Cells in the G0-G1 phase of the cell cycle, correlating with increased levels of the cell cycle inhibitor p27 as well as with reduced proliferation rates resulting in greatly diminished sensitivity of A549 cells to the late S-G2-M cell cycle arrest mediated by cisplatin. These results suggest that miR-630 may be a novel modulator of the cisplatin response in non-small cell lung cancer (71). In addition, miR-24-2 is capable of inducing apoptosis by modulating different apoptotic pathways and targeting BCL-2. Furthermore, cells overexpressing miR-24-2 are hypersensitive to DNA-damaging drugs such as cisplatin and undergo apoptotic cell death, suggesting that miR-24-2 may be involved in drug resistance in human cancer cells (MCF-7 and HeLa) (72). miR-128 is obviously expressed at a higher level in drug-resistant breast cancer samples than its expression in drug-sensitive samples. Following transfection with a precursor of miR-128 or antisense-miR-128 oligonucleotides, the chemosensitivity of MDA-MB-231 cell was upregulated (73), suggesting a direct association of miR-128 with drug resistance in cancer.

In regards to miR-216b and miR-486-5p, no study has indicated their associations with drug resistance in cancer, but they contribute to several biological processes during cancer development. It has been proven that downregulated expression of miR-216b is directly related to advanced clinical stage and lymph node metastasis. Both in vitro and in vivo assays revealed that miR-216b attenuates cell proliferation, invasion and tumor growth in nude mice (74). In addition, miR-216b, miR-186, miR-337-3p and miR-760 were found to cooperatively promote cellular senescence through the p53-p21 (Cip1/WAF1) pathway in human colorectal cancer cells (75). Given that the overexpression of miR-486-5p is closely correlated with the downregulated expression of ARHGAP5 in lung tumor tissues, which considerably inhibits lung cancer cell migration and invasion, miR-486-5p may act as a tumor suppressor contributing to the progression and metastasis of non-small cell lung cancer (76). Actually, the biological processes including cell proliferation, invasion, growth and metastasis in which miR-216b and miR-486-5p are involved are closely related to the development of drug resistance in cancers (49,77), and the annotation of biological processes also suggests that cell proliferation and growth are associated with drug resistance. All these results indicate that miR-216b and miR-486-5p may be associated with drug resistance in an indirectly way.

**Discussion**

Inferring the functional role of proteins/genes is a primary task in biology, for purposes ranging from general knowledge to drug discovery and diagnostic development (78). Protein/gene function prediction based on various genome-wide data is a potential, feasible and valuable strategy for gene function mining, and many large-scale networks of molecular interactions within the cell have made it possible to go beyond the one dimensional approach to study protein function in the context of a network (79). For example, on the basis of comprehensive bioinformatic analysis, Yin et al (77) reported that SPARCL1 and CCL21 are associated with drug resistance in ovarian cancer. Similarly, 15 TSGs associated with drug resistance in ovarian cancer were analyzed by comprehensive bioinformatics to overview the relationship of the 15 TSGs with drug resistance and to discover potential TSGs related with drug resistance (49).

Oncomine is a cancer microarray database and web-based data-mining platform aimed at facilitating the discovery from genome-wide expression analyses. Differential expression analyses comparing most major types of cancer with respective normal tissues are available for exploration. Data can be queried and visualized for a selected gene across all analyses (80). The Gene Expression Omnibus (GEO) at the National Center for Biotechnology Information (NCBI) has emerged as the leading fully public repository for gene expression data, predominantly gene expression data generated by DNA microarray technology (23). By 2006, GEO stored approximately a billion individual gene expression measurements, derived from over 100 organisms, submitted by over 1,500 laboratories, addressing a wide range of biological phenomena (81). Thus, the expression data of NEK2 in ovarian cancer tissues and drug-resistant cells retrieved from the two online databases are reliable.

GeneMANIA is a web-based database and a tool for the prediction of gene functions on the basis of
multiple networks derived from different genomic or proteomic data/sources (24). Six organisms are currently supported (Arabidopsis thaliana, Caenorhabditis elegans, Drosophila melanogaster, Mus musculus, Homo sapiens and Saccharomyces cerevisiae), and hundreds of data sets have been collected from GEO, BioGRID, Pathway Commons and 12D, as well as organism-specific functional genomics data sets (82). Thus, it is fast enough to predict gene functions with great accuracy using this software. COREMINE Medical is a product of the PubGene Company designed to be used for searching information on health, medicine and biology (25). COREMINE Medical grew out of Pubgene online tool which is a gene/protein database and a web-based tool for literature mining. Pubgene carries out automated extraction of experimental and theoretical biomedical knowledge from publicly available gene and text databases to create a gene-to-gene co-citation network for millions of named human genes by automated analysis of titles and abstracts in over 10 million MEDLINE records (83). Therefore, the functions of NEK2 predicted by these two software programs were potentially accurate.

miRNAs are a class of small (22 bp) endogenous non-coding RNAs which regulate gene expression at the post-transcriptional and translational levels (84,85). miRNA-mediated post-transcriptional gene regulation is considered as a significant regulator of many cellular processes including cell proliferation, apoptosis, differentiation, angiogenesis, invasion, metastasis and drug resistance, both physiological and pathological (86-88). miRNAs perform their functions through the regulation of their target genes, and it has been well established that miRNAs represent a class of genes with a great potential for use in diagnostics, prognosis and therapy (89). Therefore, we can predict the function of genes through the functions of the miRNAs targeting the gene. miRWalk is a comprehensive database on miRNAs, which collects predicted and validated miRNA binding sites on all known genes of the human, mouse and rat. More importantly, the miRWalk is a potential real-time database, in which the ‘Predicted Target module’ is updated every 6 months and the ‘Validated Target module’ is updated every month (26).

Taken together, based on the comprehensive bioinformatic analysis including microarray data interpretation, protein/gene-protein/gene interaction, annotation of biological processes and miRNA-mRNA interaction (Fig. 4), we revealed that the expression of NEK2 in drug-resistant ovarian cancer cells was elevated and that NEK2 directly/indirectly interacts with 20 proteins/genes which all contribute to drug resistance in ovarian and other cancers (Fig. 2). Further analysis based on the annotation of biological processes suggested that NEK2 was closely related to 14 biological processes which are involved in ovarian cancer and drug resistance (Fig. 3). Moreover, 7 of 10 miRNAs targeting NEK2 are involved in drug resistance in ovarian and other cancers (Table II). Given that NEK2 interacts with numerous proteins/genes, miRNAs and biological processes which were all found to contribute to drug resistance in ovarian and other cancers, upregulation of NEK2 in ovarian cancer and drug-resistant cancer cells may also contribute to drug-resistance.

Annotation of biological processes revealed that among the 14 biological processes annotated for NEK2, 6 were cell cycle-related (Fig. 3), and the molecular function annotation-based protein/gene-protein/gene interaction obtained similar results. These results suggest that NEK2 may participate in the regulation of drug resistance mainly through regulation of the cell cycle. In addition, microtubule-related function was another major function of NEK2 with its interacting proteins in accordance with protein/gene-protein/gene interaction (Table I), which implicated that NEK2 may also exert its drug resistant functions via microtubules. Actually, NEK2 has been reported to play important roles in both mitosis and meiosis (90) and contributes to the establishment of the microtubule-based mitotic spindle (27).

Collectively, for the first time, we report that the over-expression of NEK2 in ovarian cancers/drug-resistant cells may contribute to drug resistance through regulation of the cell cycle and microtubules. The present study provides important information for further investigation of the drug resistant-related functions of NEK2 in ovarian cancer.

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References


