

Long non-coding (lnc)RNA PAPAS overexpression inhibits tumor growth in papillary thyroid carcinoma by downregulating lncRNA HOTTIP

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Abstract. The long non-coding (lnc)RNA pre-ribosomal RNA antisense transcripts (PAPAS) can repress ribosomal RNA synthesis, although its role in human disease is currently unknown. The present study aimed to investigate the function of PAPAS in papillary thyroid carcinoma (PTC). PAPAS was downregulated, while the lncRNA HOXA transcript at the distal tip (HOTTIP) was upregulated in patients with PTC. PAPAS and HOTTIP were inversely associated in PTC. PAPAS overexpression led to the downregulation of HOTTIP in PTC cells, while PAPAS expression was not significantly affected by HOTTIP overexpression, suggesting that PAPAS was upstream of HOTTIP. PAPAS expression level decreased, while HOTTIP expression level increased with the increase of clinical staging. PAPAS overexpression led to the inhibition of cell proliferation, while HOTTIP overexpression increased proliferation of PTC cells, and HOTTIP overexpression decreased the effects of PAPAS overexpression. lncRNA PAPAS overexpression may inhibit tumor growth in papillary thyroid carcinoma by downregulating lncRNA HOTTIP.

Introduction

Papillary thyroid carcinoma (PTC) is the most frequently diagnosed type of thyroid cancer, accounting for >80% of all cases of thyroid cancer in 2010 worldwide (1). Thyroid cancer affects ~4% of all patients with cancer in 2010 worldwide (2). However, the incidence rates of this disease have increased >3-fold over the past three decades (3). Therefore, thyroid cancer is predicted to be a major type of malignancy in the

near future. PTC is usually treated with radioiodine ablation and surgery, and treatment outcomes are usually satisfactory (4). At present, >95% of patients with PTC live >5 years after diagnosis (4). However, on occasion, certain aggressive forms of PTC can invade the lymph nodes, invade local or distant tissues, or even differentiate into lethal thyroid types of cancer (5), leading to poor survival rates.

Long non-coding RNAs (lncRNAs), ncRNAs of >200 nucleotides in length, form a group of functional non-protein coding RNA transcripts (6). lncRNAs were originally considered 'noise', although accumulating evidence over the past three decades has demonstrated that they are critical determinants in both physiological processes and pathological conditions, such as the occurrence and development of cancer (7-9). Therefore, regulation of lncRNA expression may provide new insights into the treatment or even prevention of cancer (10). However, the clinical application of lncRNAs is limited by the unknown function of the majority of lncRNAs. It has been previously reported that pre-ribosomal RNA antisense transcript (PAPAS) is able to repress ribosomal RNA (rRNA) synthesis (11). Our previous transcriptome analysis (12) revealed that PAPAS was downregulated in PTC tissues, and its expression levels were inversely associated with lncRNA HOXA transcript at the distal tip (HOTTIP), which is an oncogenic lncRNA in PTC (12). The present study assessed the involvement of PAPAS in PTC and investigated its association with HOTTIP.

Materials and methods

Patients. The present study enrolled 58 patients (21 males and 37 females) with PTC from the Qingdao Municipal Hospital (Qingdao, China), between January 2015 and January 2018. The mean age of the patients was 45.2±5.2 years, (age range; 32-58 years). The inclusion criteria for the patients were as follows: i) Patients with PTC that had been diagnosed by imaging and histopathological examinations; ii) had no previous history of malignancy; iii) the patient was diagnosed with PTC, but had received no previous therapy; and iv) had normal thyroid function. The exclusion criteria for the patients were as follows: i) Patients had multiple clinical disorders

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diagnosed in addition to PTC; and ii) had received any form of treatment. Based on the American Joint Committee on Cancer system (13), there were 15, 16, 13 and 14 cases at stage I, II, III and IV, respectively, in the present study. There were 26 cases at high grade and 32 cases at low grade (14). A total of 20 patients with thyroid goiter (8 males and 12 females; 34-56 years; 45.8±5.8 years) were also selected during the same time period at the aforementioned hospital. All patients provided written informed consent prior to the start of the study. The present study was approved by the Ethics Committee of Qingdao Municipal Hospital.

Specimens and cell lines. Thyroid biopsy was performed on all patients in order to collect specimens of both tumor tissues and adjacent (within 2 cm from the tumor tissues) healthy tissues. All specimens were confirmed by three experienced pathologists at the aforementioned hospital (blinded). Thyroid biopsies were also obtained from the 20 patients with thyroid goiter. All thyroid biopsies from patients with thyroid goiter were stored in a liquid nitrogen sink at specimen library of the Qingdao Municipal Hospital.

IHH-4 and HTH-83 PTC cell lines and Nthy-ori 3-1 thyroid follicular epithelial cell line were used in the present study to perform all the *in vitro* cell experiments. Cells were bought from the American Type Culture Collection. DMEM supplemented with 100 mg/ml penicillin G, 10% FBS (Sigma-Aldrich; Merck KGaA), and 100 U/ml streptomycin (Invitrogen; Thermo Fisher Scientific, Inc.) was used to culture cells in an incubator at 37°C with 5% CO₂ to reach about 80% confluence

Total RNA extraction and reverse transcription-quantitative PCR (RT-qPCR). Total RNA in both tissue specimens and *in vitro* cells were extracted using TRIzol[®] reagent (Invitrogen; Thermo Fisher Scientific, Inc.) according to manufacturer's instructions. Tumor and adjacent healthy tissues were frozen in liquid nitrogen (-196°C) prior to the addition of the TRIzol reagent. Following RNA extraction, SuperScript III Reverse Transcriptase kit (Thermo Fisher Scientific, Inc.) was used to perform the reverse transcription to synthesize cDNA through the following thermal conditions: 55°C for 30 min and 80°C for 10 min. All PCR reaction mixtures were prepared using the SYBR Green Master mix (Bio-Rad Laboratories, Inc.). 18S rRNA was used as the endogenous control. The 2^{-ΔΔC_q} method (15) was used to process the data. The primer sequences used were as follows: PAPAS forward, 5'-ATGGGGCCA AGATTGTGTCT-3' and reverse, 5'-AGACACAATCTTGCC CCCAT-3'; HOTTIP forward, 5'-AAGGCGGTTTTACATACT GGTC-3' and reverse, 5'-TAGCACCTGTAGTTGCCCATC CC-3'; 18S rRNA forward, 5'-GGCCCTGTAATTGGAATG AG-3' and reverse, 5'-CCAAGATCCAACACTACGAGCTT-3'. The thermocycling conditions were as follows: 95°C for 5 min, followed by 40 cycles of 95°C for 10 sec and 55°C for 40 sec.

Vector constructions and cell transfection. pcDNA3.1 vectors expressing PAPAS and HOTTIP were constructed by Sangon Biotech Co., Ltd. An empty vector (pcDNA3.1) was used as negative control (NC). NC small interfering (si)RNA (5'-UUC UCCGAACGUGUCACGUUU-3') and HOTTIP siRNA (5'-GC CGCCGUGUCCACCGGCAGCU-3') were also obtained from

Sangon Biotech Co., Ltd. IHH-4 and HTH-83 PTC cells were harvested once they reached 70-80% confluence, and Lipofectamine 2000 (Thermo Fisher Scientific, Inc.) was used to transfect 10 nM plasmid or 40 nM siRNA into 10⁶ cells. The cells were harvested 24 h post-transfection prior to subsequent experiments.

Cell proliferation analysis. The effects of transfections on cell proliferation were analyzed using a Cell Counting Kit-8 (CCK-8; Sigma-Aldrich; Merck KGaA) assay according to manufacturer's instructions. Single cell suspensions were prepared using DMEM and cell concentration was diluted to 4x10³ cells/ml. Cell culture was then performed using a 96-well plates (100 μl per well). The plates were incubated in an incubator at 37°C with 5% CO₂, followed by the addition of 10 μl CCK-8 solution every 24 h for 96 h. Cells were then cultured for an additional 4 h, followed by the addition of 10 μl DMSO. Finally, optical density values at 450 nm were measured to analyze cell proliferation.

Statistical analysis. All statistical analysis was performed using GraphPad Prism 6 software (GraphPad Software, Inc.). Experiments were repeated in triplicate, and the results are presented as the mean ± SD. Differences were assessed using one-way ANOVA followed by Tukey's test. Associations between the expression levels of two genes were analyzed by linear regression analysis. P<0.05 was considered to indicate a statistically significant difference.

Results

Expression levels of PAPAS and HOTTIP are altered in tumor tissues of patients with PTC. Differential gene expression provides strong evidence for the involvement of certain genes in various pathological processes. Therefore, the present study investigated the expression of PAPAS and HOTTIP in both tumor and adjacent healthy tissues via RT-qPCR. Compared with the adjacent healthy tissues, expression levels of PAPAS were significantly lower in tumor tissues and thyroid biopsies from patients with thyroid goiter (P<0.05; Fig. 1A). By contrast, lncRNA HOTTIP levels were upregulated in tumor tissues and thyroid biopsies from thyroid goiter patients than in adjacent healthy tissues (P<0.05; Fig. 1B). However, no significant differences were observed between tumor tissues and thyroid biopsies from patients with thyroid goiter.

PAPAS and HOTTIP are inversely associated in PTC. Correlations between PAPAS and HOTTIP were analyzed by linear regression analysis. It was observed that PAPAS and HOTTIP were inversely and significantly associated in tumor tissues (P<0.0001; Fig. 2A). However, the expression levels of PAPAS and HOTTIP in adjacent healthy tissues were not significantly associated (P=0.1420; Fig. 2B), indicating that PAPAS and HOTTIP were associated specifically under pathological conditions.

PAPAS is an upstream negative regulator of HOTTIP in PTC cell lines. Expression levels of PAPAS and HOTTIP in IHH-4, HTH-83 and Nthy-ori 3-1 cell lines were measured by RT-qPCR. Compared with Nthy-ori 3-1 cells, PAPAS was

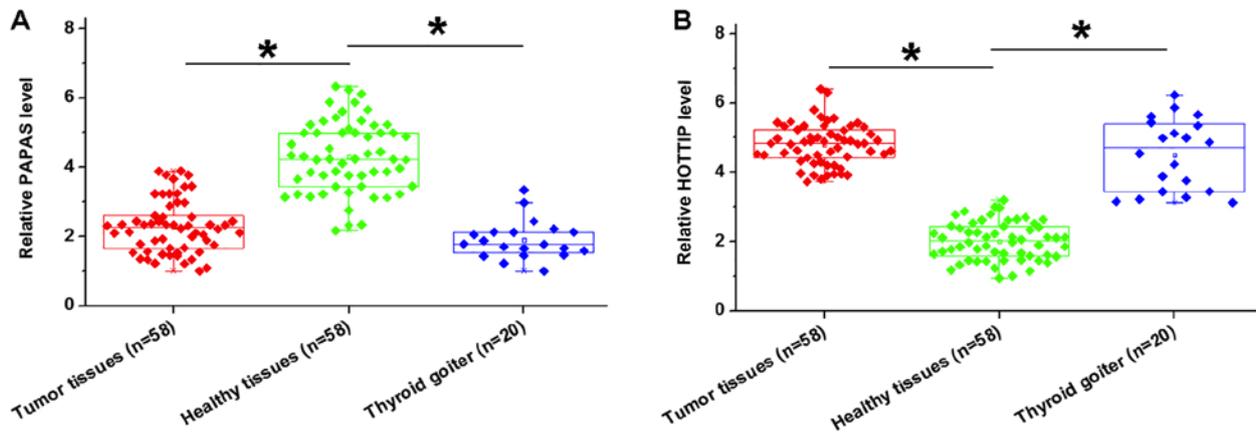


Figure 1. Expression levels of PAPAS and HOTTIP are altered in the tumor tissues of patients with PTC. Reverse transcription-quantitative PCR revealed that (A) PAPAS was downregulated, while (B) long non-coding RNA HOTTIP was upregulated in tumor tissues and thyroid biopsies from patients with thyroid goiter compared with adjacent healthy tissues of patients with papillary thyroid carcinoma. No significant differences were observed between thyroid biopsies from patients with thyroid goiter and thyroid biopsies from patients with PTC. * $P < 0.05$ vs. healthy tissues. PAPAS, pre-ribosomal RNA antisense transcripts; HOTTIP, HOXA transcript at the distal tip; PTC, papillary thyroid carcinoma.

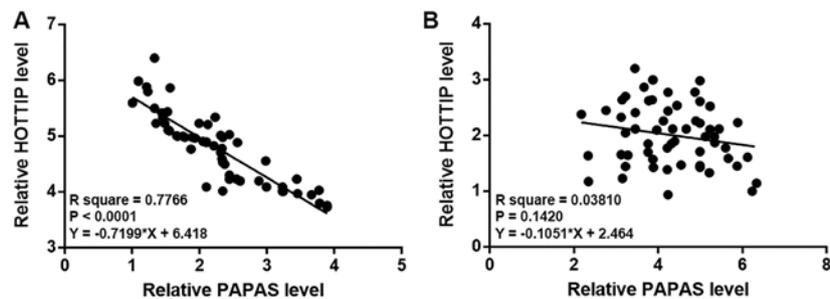


Figure 2. PAPAS and HOTTIP are inversely associated in papillary thyroid carcinoma. The linear regression analysis revealed that PAPAS and HOTTIP were inversely associated in (A) tumor tissues, but (B) not in adjacent healthy tissues. PAPAS, pre-ribosomal RNA antisense transcripts; HOTTIP, HOXA transcript at the distal tip.

downregulated ($P < 0.05$; Fig. S1A) and HOTTIP was upregulated ($P < 0.055$; Fig. S1B) in IHH-4 and HTH-83 cells. In order to further investigate the interactions between PAPAS and HOTTIP, PAPAS and HOTTIP vectors were transfected into the IHH-4 and HTH-83 cell lines. The RT-qPCR results revealed that PAPAS and HOTTIP were overexpressed at 24 h after transfection in the IHH-4 and HTH-83 cell lines ($P < 0.05$; Fig. 3A). Compared with the untransfected (C) and NC groups, cells overexpressing PAPAS exhibited significantly downregulated expression levels of HOTTIP ($P < 0.05$; Fig. 3B). However, the expression level of PAPAS was not significantly altered in cells overexpressing HOTTIP (Fig. 3C). HOTTIP siRNA silencing was achieved in both IHH-4 and HTH-83 cells ($P < 0.05$; Fig. S2A). Compared with the C and NC groups, HOTTIP siRNA silencing failed to significantly affect the expression of PAPAS in these cells (Fig. S2B).

Expression levels of PAPAS and HOTTIP are affected by clinical stage. Comparison among patients with different clinical stages revealed that expression levels of PAPAS decreased significantly along with the increase in clinical staging ($P < 0.05$; Fig. 4A). In contrast, HOTTIP expression level increased significantly with the increase of clinical staging ($P < 0.05$; Fig. 4B). The present results indicated that PAPAS and HOTTIP are involved in the progression of PTC.

PAPAS regulates PTC cell proliferation through HOTTIP. Compared with C and NC groups, PAPAS overexpression led to decreased cell proliferation, whereas HOTTIP overexpression increased the proliferation of PTC cells, and HOTTIP overexpression attenuated the effects of PAPAS overexpression in both IHH-4 and HTH-83 cells ($P < 0.05$; Fig. 5).

Discussion

Although PAPAS has been reported to play a critical role in rRNA synthesis (11), its role in human disease remains unknown. The present study revealed that PAPAS was downregulated in PTC tissues and PTC cell lines, and played a tumor suppressive role in this disease. However, HOTTIP was upregulated in both PTC tissues and PTC cell lines. In addition, the role of PAPAS in PTC was found to be mediated by the downregulation of oncogenic HOTTIP.

Accumulating evidence demonstrated that HOTTIP promotes the development and progression of different types of cancer, such as pancreatic cancer and esophageal cancer (16,17). HOTTIP is involved in cancer development and progression primarily by regulating cancer cell behaviors via its interactions with downstream tumor suppression of oncogenic pathways, such as HOXA13 and the epithelial-to-mesenchymal transition pathways (16,17). In a recent study, Yuan *et al* (12)

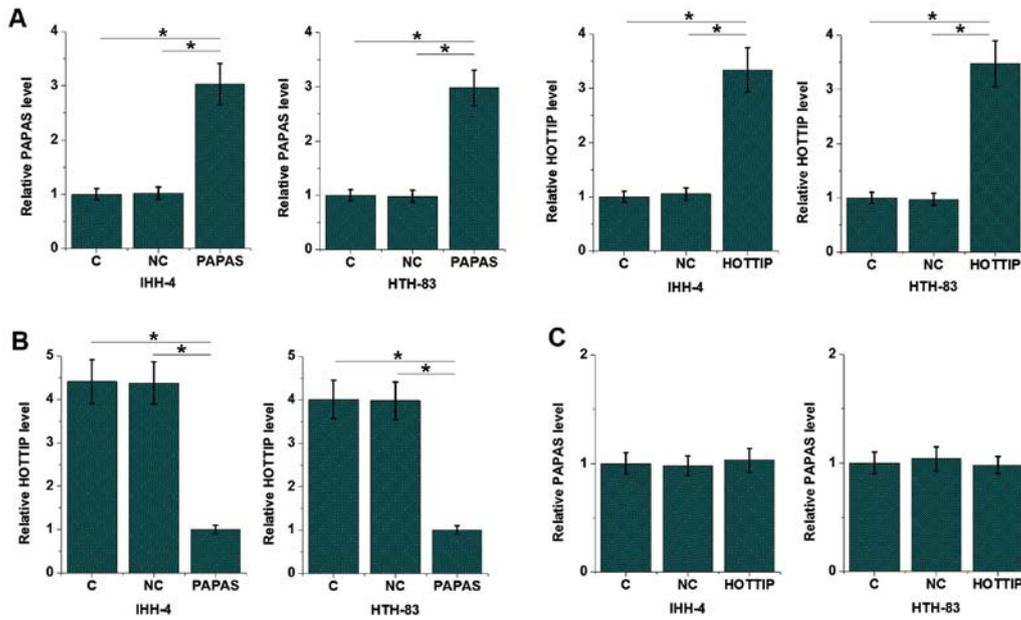


Figure 3. PAPAS is an upstream negative regulator of HOTTIP in IHH-4 and HTH-83 cell lines. (A) PAPAS and HOTTIP were upregulated 24 h after transfection. Compared with the C and NC groups, (B) PAPAS overexpression led to the downregulation of HOTTIP in IHH-4 and HTH-83 cells, while (C) HOTTIP overexpression failed to significantly affect PAPAS expression. *P<0.05. PAPAS, pre-ribosomal RNA antisense transcripts; HOTTIP, HOXA transcript at the distal tip; C, untransfected control; NC, negative control.

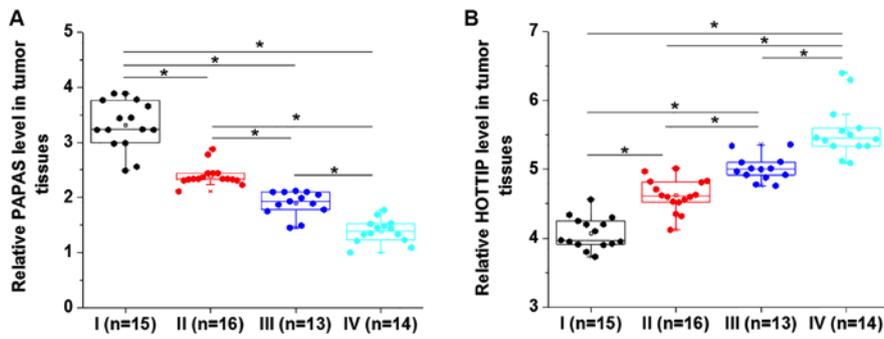


Figure 4. Expression levels of PAPAS and HOTTIP are affected by clinical stage. (A) PAPAS expression level decreased, while (B) HOTTIP expression level increased with the increase of clinical staging. *P<0.05. PAPAS, pre-ribosomal RNA antisense transcripts; HOTTIP, HOXA transcript at the distal tip.

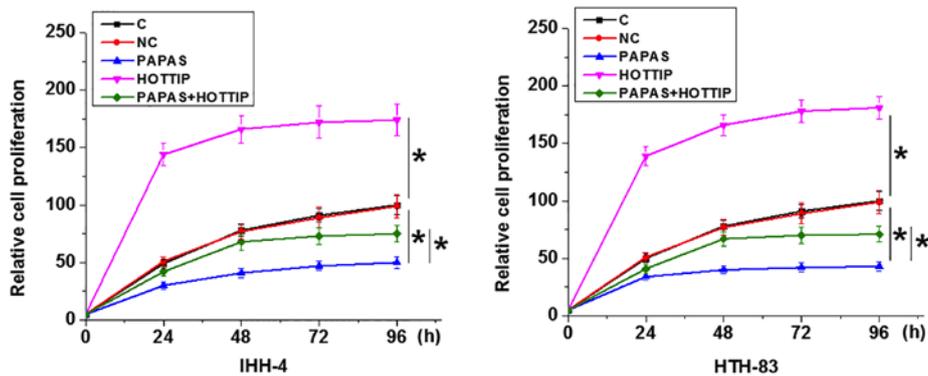


Figure 5. PAPAS regulates IHH-4 and HTH-83 cell proliferation via HOTTIP. The *in vitro* cell proliferation assay data revealed that PAPAS overexpression led to decreased cell proliferation, while HOTTIP overexpression led to increased proliferation of papillary thyroid carcinoma cells, and HOTTIP overexpression attenuated the effects of PAPAS overexpression. *P<0.05. PAPAS, pre-ribosomal RNA antisense transcripts; HOTTIP, HOXA transcript at the distal tip; C, control; NC, negative control.

reported that HOTTIP was upregulated in PTC and was able to promote cancer cell proliferation, invasion and migration

through the downregulation of microRNA-637, which inhibits cancer development (12). Consistent with previous results (12),

the present study observed the upregulated HOTTIP in PTC tissues, and the overexpression of HOTTIP led to significantly increased cancer cell proliferation. Therefore, the present study further confirmed the oncogenic role of HOTTIP in PTC.

To the best of our knowledge, the present study is the first to report the tumor suppressive role of PAPAS in PTC. lncRNAs participate in cancer biology, primarily by interacting with downstream oncogenes or tumor suppressors (18,19). Notably, the present study indicated that PAPAS could serve a role in cancer development acting as an upstream inhibitor of HOTTIP in PTC. However, the interaction between PAPAS and HOTTIP was found to be likely indirect, due to the lack of association between the expression levels of these two lncRNAs in tumor adjacent healthy tissues. HOTTIP overexpression only partially attenuated the inhibitory effects of PAPAS overexpression on cancer cell proliferation. Therefore, it is likely that PAPAS may interact with multiple downstream effectors to achieve a fine regulation of PTC cell behaviors.

However, the present study is limited by the small sample size. In addition, studies with *in vivo* animal models are required in order to further validate the conclusions from the present study. However, the present study provided evidence of the existence of the interactions between different lncRNAs in cancer biology. The present results provide new insights into cancer biology. However, the molecular mechanisms underlying the interactions between lncRNAs are unclear.

PAPAS and HOTTIP were also dysregulated in thyroid goiter. Therefore, the interaction between PAPAS and HOTTIP is unlikely to be cancer-specific. Future studies should aim to investigate the involvement of PAPAS and HOTTIP in other types of thyroid disease.

In conclusion, PAPAS was downregulated in PTC and may inhibit PTC cell proliferation by downregulating HOTTIP.

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Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Authors' contributions

JX and JL designed the experiments. JX and ZB performed experiments. GX and YG analyzed the data. JL drafted the manuscript. All authors approved the manuscript.

Ethics approval and consent to participate

The present study was approved by Ethics Committee of Qingdao Municipal Hospital (Qingdao, China). All patients provided written informed consent prior to the study start.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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