

Levels of acute inflammatory biomarkers in advanced prostate cancer patients with α_2 -macroglobulin deficiency

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Abstract. C-reactive protein (CRP), serum amyloid A (SAA), interleukin-6 (IL-6), α_1 -antitrypsin (α_1 AT), α_1 -acid glycoprotein (α_1 AG) and ceruloplasmin (CP) are acute inflammatory biomarkers that increase in various conditions including infection, inflammation, malignancy and tissue disturbance. In contrast, α_2 -macroglobulin (α_2 M) is involved in inflammation through its function as a carrier protein of IL-6. We had previously reported on advanced prostate cancer (PCa) patients with multiple distant bone metastases in whom serum α_2 M levels were markedly decreased (α_2 M deficiency). However, the relationship between serum levels of α_2 M and acute inflammatory biomarkers in PCa patients with or without α_2 M deficiency has not been demonstrated. In the present study, we examined serum levels of CRP, SAA, IL-6, α_1 AT, α_1 AG and CP in PCa patients with or without α_2 M deficiency to establish clinical significance and changes in these biomarkers during PCa disease progression. We found that upon addition of recombinant IL-6 (rIL-6) to serum from PCa patients with α_2 M deficiency, since a function of α_2 M is to bind and stabilize IL-6, the α_2 M-IL-6 complex and free endogenous IL-6 were not detectable. Serum levels of the α_2 M-independent markers, α_1 AT, α_1 AG and CP, in all PCa patients regardless of α_2 M deficiency were significantly higher than in healthy controls, but those of the α_2 M-dependent molecules, CRP, SAA and IL-6, were not increased in PCa patients with α_2 M deficiency. Therefore, quantitation of both α_2 M-dependent (CRP, SAA and IL-6) and α_2 M-independent (α_1 AT, α_1 AG and CP) acute inflammatory biomarkers in advanced PCa patients may be an auxiliary indicator, together with prostate-specific antigen (PSA), to monitor PCa disease progression.

Introduction

α_2 -macroglobulin (α_2 M) is the most abundant proteinase inhibitor in the blood and inhibits the activity of various proteases through direct interaction. Moreover, a major biological function of α_2 M is as a carrier protein for interleukin-6 (IL-6) or growth hormone (1,2). Therefore, α_2 M is involved in coagulation, fibrinolytic activity and inflammatory reaction *in vivo*. Serum α_2 M levels generally increase in various disorders, such as nephrotic syndrome, inflammatory diseases and malignancy. On the other hand, it is also known that serum α_2 M levels decrease in very few disorders such as disseminated intravascular coagulation (DIC) and hematologic malignancy. We had previously reported on advanced prostate cancer (PCa) with multiple distant bone metastases, in whom serum α_2 M levels were markedly decreased to <20 mg/dl (α_2 M deficiency), while serum prostate-specific antigen (PSA) levels were greatly increased (3,4). These cases were not complicated by DIC.

IL-6, C-reactive protein (CRP) and serum amyloid A (SAA) are widely used as acute inflammatory biomarkers in various conditions such as infection, inflammation, malignancy and tissue disturbance (5-7). IL-6 is a multi-functional cytokine that regulates the production of CRP and SAA in liver cells in various conditions (8,9). CRP is most widely used as a sensitive inflammatory biomarker in routine clinical examination. SAA exists as a high density lipoprotein (HDL)-complex with a molecular weight of 200-400 kDa, which is generally increased in patients with viral infection or in corticosteroid-treated patients in contrast to CRP (6,10).

α_1 -antitrypsin (α_1 AT) is a plasma glycoprotein, which inhibits the activity of proteases, such as trypsin and elastase, derived from macrophage or pancreas cells. α_1 -acid glycoprotein (α_1 AG) is also a plasma glycoprotein that inhibits the activity of progesterone. Ceruloplasmin (CP) is a copper-binding protein and is involved in metabolism of copper or iron.

It is thought that α_2 M is involved in inflammatory reaction through its function as a carrier protein of IL-6 (1). Furthermore, it has been suggested that the concentration of α_2 M affects the levels of IL-6, CRP and SAA produced by liver cells through its regulation of IL-6. However, the relationship between serum levels of α_2 M and acute inflammatory biomarkers in PCa

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patients with or without $\alpha 2M$ deficiency has not been demonstrated. Therefore, we quantified serum levels of CRP, SAA, IL-6, $\alpha 1AT$, $\alpha 1AG$ and CP in PCa patients with or without $\alpha 2M$ deficiency to establish the clinical significance and changes of these acute inflammatory biomarkers during PCa disease progression. Although this study includes only a limited number of PCa patients, it is the first report to investigate the clinical significance and changes of acute inflammatory biomarkers in PCa patients with regard to $\alpha 2M$ deficiency.

Materials and methods

Subjects. Forty-three untreated adult men participated in this study, of whom 10 were healthy controls (mean age 62.8 years, range 52-70) and 33 were diagnosed with prostatic disease at the Kitasato University Hospital. The thirty-three patients had PCa at stage M1b (mean age 67.8 years, range 55-79) without inflammatory disorders. $\alpha 2M$ deficiency was defined as serum $\alpha 2M$ levels <20 mg/dl and there was no precipitation line of $\alpha 2M$ on immunoelectrophoresis with anti-whole human serum and antiserum to $\alpha 2M$ in routine examination. These patients also had markedly increased serum PSA levels and multiple distant bone metastases. The histological diagnoses in the patients were confirmed by six-sextant biopsy and/or transurethral resection in all cases. PCa was staged clinically following the TNM classification (11). Briefly, stage T1 is defined as tumor not clinically recognizable and identifiable only by histological examination of prostatic tissue. Stage T2 tumors are palpable but confined within the prostate. Stage T3 tumors are palpable and extend through the prostatic capsule with unilateral or bilateral extension. The M1 stage is defined by the presence of distant metastasis and M1b by bone metastasis. Serum samples were obtained from these patients and stored at $-80^{\circ}C$ until use.

Ethical approval. The study was conducted in accordance with the Declaration of Helsinki. This study had no influence on the management of patients, and informed consent was obtained from all subjects.

Acute inflammatory biomarkers. Measurement of $\alpha 2M$, $\alpha 1AT$, $\alpha 1AG$ and CP levels in serum was performed by laser nephelometry using a Behring Nephelometer Analyzer (Behring Diagnostics, Westwood, MA). CRP and SAA levels in serum were measured by latex nephelometry using the LX-M (Eiken Chemical Co., Japan). IL-6 levels in serum were measured by a specific luminescence sandwich-type enzyme-linked immunosorbent assay (ELISA), using a previously described method with modifications (12). The assay employed 2 monoclonal anti-human IL-6 antibodies (clones IG61 and IG67; Toray, Tokyo, Japan) and alkaline phosphatase with the chemiluminescent substrate Lumi-phos (Lumigen Inc., Detroit, MI) and was sensitive to 0.5 pg/ml.

Bone scintigraphy. Bone metastasis in PCa patients was diagnosed by bone scintigraphy using ^{99m}Tc -labeled methylene diphosphonate (^{99m}Tc -MDP).

Western blotting. Binding assays for purified $\alpha 2M$ (Protogen AG, Laufelfingen, Switzerland) and recombinant IL-6 (rIL-6)

(Genzyme, Cambridge, MA) were analyzed by Western blotting with 5-15% SDS-polyacrylamide gel electrophoresis and staining with anti-IL-6 sera (Dako, Glostrup, Denmark) according to the Laemmli method (13). $\alpha 2M$ and IL-6 of 1:1 molar concentrations were mixed and incubated for 20 min at $37^{\circ}C$.

High performance liquid chromatography. We analyzed the IL-6 elution profiles of a mixture of purified $\alpha 2M$ (50 μg) and rIL-6 (2 μg) or serum (10 μl) from patients with $\alpha 2M$ deficiency mixed with rIL-6 (2 μg) by high performance liquid chromatography (HPLC). Fractionation of serum samples was performed by HPLC using a TSKG-3000 SWXL (Tosoh, Tokyo, Japan) column, and IL-6 level in each fraction was measured. Flow rate was 1 ml/min and fractions of 0.5 ml volume were collected. The column was calibrated using 20 mM phosphate buffer (pH 7.5).

Statistical analysis. The Wilcoxon and the Mann-Whitney U-tests were used for statistical analyses, and $p < 0.05$ was considered statistically significant.

Results

Concentrations of $\alpha 2M$, CRP, SAA, IL-6, $\alpha 1AT$, $\alpha 1AG$ and CP in serum. Table I shows the concentrations (mean \pm SD) of CRP, SAA, IL-6, $\alpha 1AT$, $\alpha 1AG$ and CP in the sera of healthy controls and PCa patients with or without $\alpha 2M$ deficiency. $\alpha 2M$ deficiency was defined as serum $\alpha 2M$ levels <20 mg/dl. The serum levels of CRP (Fig. 1), SAA (Fig. 2) and IL-6 (Fig. 3) in PCa patients without $\alpha 2M$ deficiency were significantly higher than in healthy controls and PCa patients with $\alpha 2M$ deficiency (all $p < 0.01$). On the other hand, serum levels of $\alpha 1AT$ (Fig. 4), $\alpha 1AG$ (Fig. 5) and CP (Fig. 6) in PCa patients with or without $\alpha 2M$ deficiency were significantly higher than in healthy controls ($p < 0.05$). However, there was no significant difference between PCa patients with and without $\alpha 2M$ deficiency.

Western blotting for $\alpha 2M$ and IL-6 complex. To verify the detection and size of the $\alpha 2M$ -IL-6 complex, purified $\alpha 2M$ and rIL-6 were mixed, and the complex was identified using Western blotting as a band of ~ 800 kDa (Fig. 7).

IL-6 elution profiles of mixture of purified $\alpha 2M$ and rIL-6, and serum from patients with $\alpha 2M$ deficiency mixed with rIL-6. Fig. 8 shows the IL-6 elution profiles of a mixture of purified $\alpha 2M$ and rIL-6 (dots) and of serum from patients with $\alpha 2M$ deficiency mixed with rIL-6 (triangles) by HPLC. Three peaks were identified from the mixture of purified $\alpha 2M$ and rIL-6: the first peak at 800 kDa corresponding to the $\alpha 2M$ -IL-6 complex; a second peak at ~ 50 kDa corresponding to a polymer of IL-6; and a third peak at ~ 20 kDa representing free IL-6. When the serum from PCa patients with $\alpha 2M$ deficiency was mixed with rIL-6 the first and third peaks corresponding to $\alpha 2M$ -IL-6 complex and free IL-6, respectively, were undetectable.

Discussion

CRP, SAA, IL-6, $\alpha 1AT$, $\alpha 1AG$ and CP are known as acute inflammatory biomarkers that increase in various conditions such as infection, inflammation, malignancy and tissue

Table I. Concentration (mean \pm SD) of α_2 -macroglobulin (α_2 M), C-reactive protein (CRP), serum amyloid A (SAA), interleukin-6 (IL-6), α_1 anti-chymotripsin (α_1 AT), α_1 acid glycoprotein (α_1 AG) and ceruloplasmin (CP) in sera of healthy controls and advanced prostate cancer (PCa) patients with or without α_2 M deficiency.

	Healthy controls (n=10)	Advanced PCa patients	
		without α_2 M deficiency (n=23)	with α_2 M deficiency (n=10)
α_2 M (mg/dl)	151.7 \pm 25.30	252.45 \pm 125.07 ^a	12.75 \pm 5.60 ^b
CRP (μ g/dl)	109.58 \pm 89.66	1534.6 \pm 1482.8 ^a	173 \pm 90.3 ^b
SAA (μ g/ml)	9.96 \pm 4.16	54.7 \pm 44.5 ^a	2.15 \pm 1.68 ^b
IL-6 (pg/ml)	7.58 \pm 6.0	46.50 \pm 40.65 ^a	3.38 \pm 2.61 ^b
α_1 AT (mg/dl)	122 \pm 28.0	293.76 \pm 68.51 ^c	305.71 \pm 71.30 ^c
α_1 AG (mg/dl)	67.5 \pm 25.5	102 \pm 39.36 ^c	100.8 \pm 38.90 ^c
CP (mg/dl)	29.0 \pm 8.0	37.74 \pm 10.93 ^c	37.6 \pm 10.89 ^c

^ap<0.01, healthy controls vs. PCa patients without α_2 M deficiency; ^bp<0.01, PCa patients without α_2 M deficiency vs. PCa patients with α_2 M deficiency; ^cp<0.05, healthy controls vs. PCa patients with or without α_2 M deficiency.

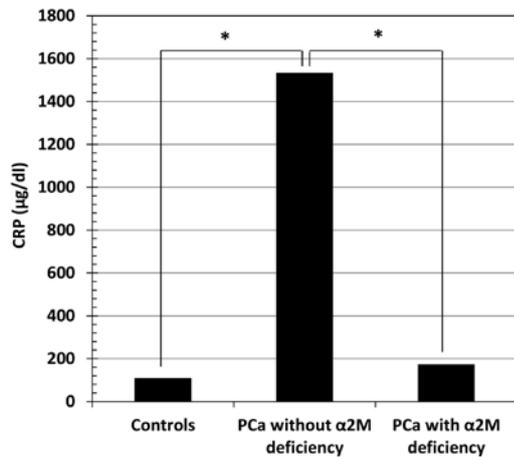


Figure 1. Serum CRP levels in healthy controls and advanced PCa patients with or without α_2 M deficiency. Serum CRP levels were significantly increased in PCa patients without α_2 M deficiency as compared with healthy controls and PCa patients with α_2 M deficiency. PCa, prostate cancer; *p<0.01.

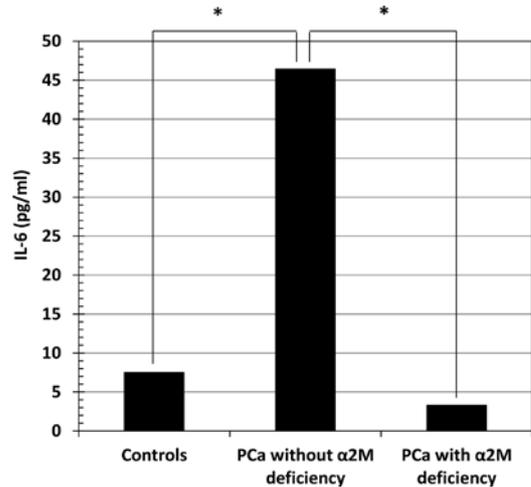


Figure 3. Serum IL-6 levels in healthy controls and advanced PCa patients with or without α_2 M deficiency. Serum IL-6 levels were significantly increased in PCa patients without α_2 M deficiency as compared with healthy controls and PCa patients with α_2 M deficiency. PCa, prostate cancer; *p<0.01.

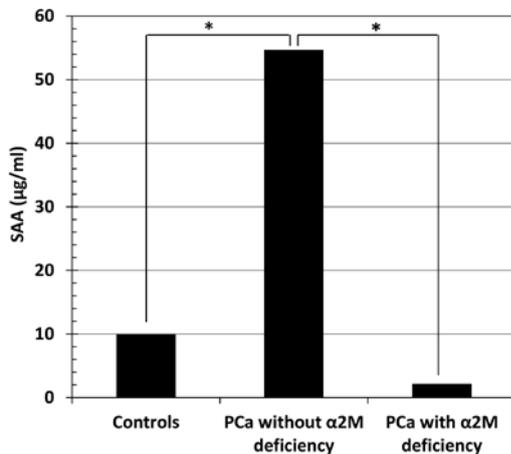


Figure 2. Serum SAA levels in healthy controls and advanced PCa patients with or without α_2 M deficiency. Serum SAA levels were significantly increased in PCa patients without α_2 M deficiency as compared with healthy controls and PCa patients with α_2 M deficiency. PCa, prostate cancer; *p<0.01.

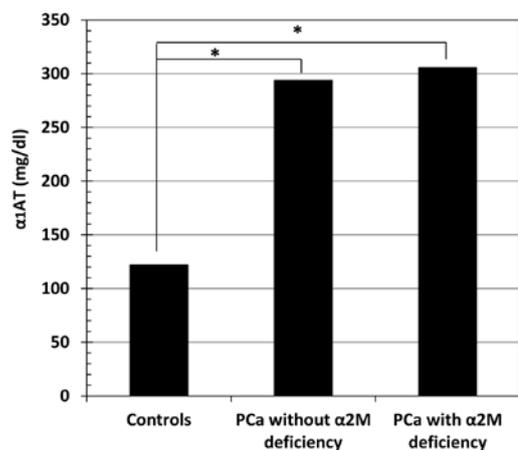


Figure 4. Serum α_1 AT levels in healthy controls and advanced PCa patients with or without α_2 M deficiency. Serum α_1 AT levels were significantly increased in PCa patients with or without α_2 M deficiency as compared with healthy controls. PCa, prostate cancer; *p<0.05.

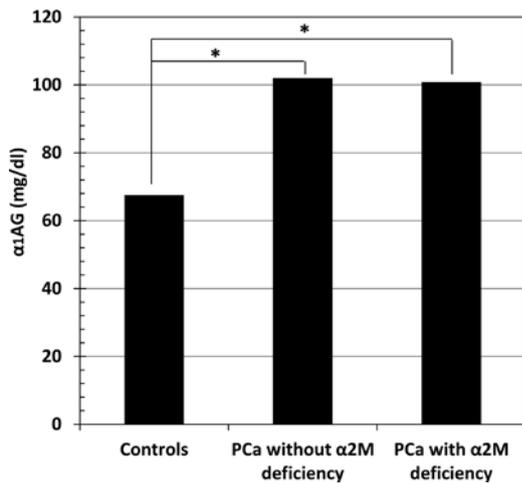


Figure 5. Serum α 1AG levels in healthy controls and advanced PCa patients with or without α 2M deficiency. Serum α 1AG levels were significantly increased in PCa patients with or without α 2M deficiency as compared with healthy controls. PCa, prostate cancer; * p <0.05.

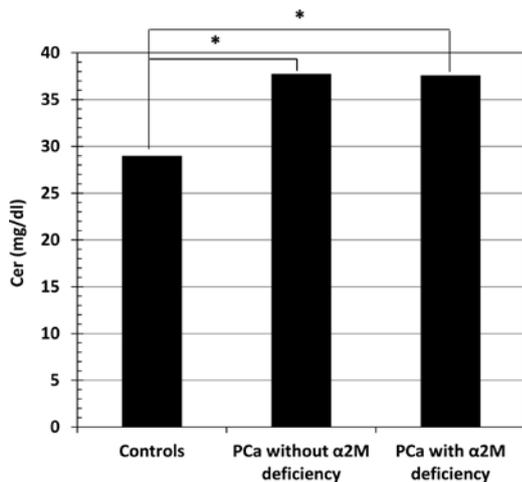


Figure 6. Serum CP levels in healthy controls and advanced PCa patients with or without α 2M deficiency. Serum CP levels were significantly increased in PCa patients with or without α 2M deficiency as compared with healthy controls. PCa, prostate cancer; * p <0.05.

disturbance (5-7,14). On the other hand, α 2M is involved in the inflammatory reaction through its function as a carrier protein of IL-6 which promotes the production of acute inflammatory biomarkers in liver cells (1). Therefore, it has been suggested that the concentration of α 2M affects the levels of IL-6, CRP and SAA produced by the liver cells through its regulation of IL-6. We had previously reported on advanced PCa patients with multiple distant bone metastases in whom serum α 2M levels were markedly decreased to <20 mg/dl (α 2M deficiency) (3,4). However, the relationship between serum levels of α 2M and acute inflammatory biomarkers in PCa patients with or without α 2M deficiency has not been demonstrated. In this study, we examined serum levels of CRP, SAA, IL-6, α 1AT, α 1AG and CP in PCa patients with or without α 2M deficiency to establish clinical significance of these biomarkers during PCa disease progression. This is the first report that analyzed

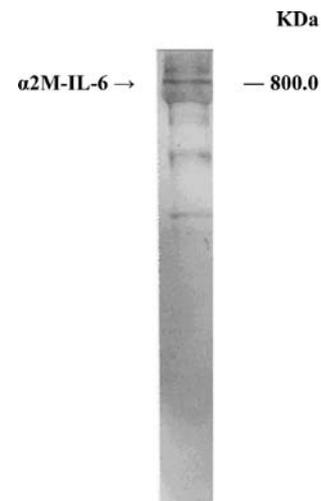


Figure 7. Western blotting for α 2M and IL-6 complex. The band at ~800 kDa corresponds to the complex between purified α 2M and recombinant IL-6.

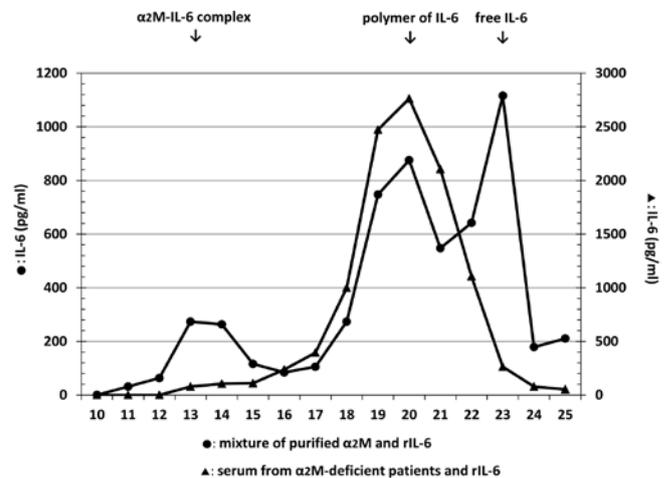


Figure 8. IL-6 elution profiles of a mixture of α 2M and recombinant IL-6 (rIL-6), and serum from patients with α 2M deficiency mixed with rIL-6. Using HPLC, the mixture of purified α 2M and rIL-6 generated three peaks: the first peak at 800 kDa corresponding to a α 2M-IL6 complex; a second peak at ~50 kDa corresponding to a polymer of IL-6; and a third peak at ~20 kDa representing free IL-6.

the changes of acute inflammatory biomarkers in PCa patients with or without α 2M deficiency.

CRP is a plasma protein produced in liver cells by cytokine stimulation, mainly IL-6, but also interleukin-1 β (IL-1 β) and tumor necrosis factor- α (TNF- α) secreted from macrophages (8,9). Serum CRP levels are increased in various conditions including infection, inflammation and tissue disturbance such as malignancy or myocardial infarction, but rarely in viral infection, multiple myeloma and non-active systemic lupus erythematosus (6,15,16). It has been reported that CRP and SAA may be important prognostic markers for long-term survival in breast cancer patients (17). Fujikawa *et al* showed that elevated serum CRP levels are associated with tumor progression and poor prognosis of esophageal cancer (7). In recent years, the determination of high sensitivity CRP (hs-CRP) is possible due to the wide use of low concentration range measurements in

routine clinical examination. It has been reported that hs-CRP reflects the degree of localized vascular inflammation and is a useful prognostic marker of cardiovascular events (18,19).

SAA is a plasma protein produced in liver cells by cytokine stimulation mainly IL-1 β , as well as IL-6 and TNF- α , and it exists as a high density lipoprotein (HDL)-complex with a molecular weight of 200-400 kDa. SAA is generally increased in patients with viral infection and corticosteroid-treated patients, a characteristic that differs from CRP (6,10). It has been hypothesized that the degree of change in serum SAA levels is larger as compared to CRP in various conditions such as infection, inflammation, malignancy and tissue disturbance, and IL-1 β stimulation of SAA production is hard to suppress by corticosteroid (10). Cocco *et al* showed that SAA may be a novel biomarker for endometrial cancer to monitor disease recurrence and response to therapy (20). It is also reported that extremely elevated plasma levels of CRP and SAA are high risk factors for development of gastric cancer (21).

IL-6 is the most sensitive inflammatory cytokine which promotes the production of both CRP and SAA in liver cells, similar to IL-1 β and TNF- α . It has been shown that serum IL-6 and CRP levels may be useful in the differentiation between diagnosis of pancreatic cancer and chronic pancreatitis (22). Kim *et al* reported that preoperative serum IL-6 and CRP levels may be markers of tumor invasion, lymph node metastasis and TNM stage (23). In the present study, CRP, SAA and IL-6 levels in serum were significantly increased in PCa patients without α 2M deficiency as compared with healthy controls and PCa patients with α 2M deficiency.

To further study the relationship between IL-6 and α 2M, we demonstrated and confirmed here that IL-6 can easily complex with α 2M *in vitro* as detected by immunoblotting. Furthermore, using HPLC, we showed that the IL-6 elution profiles of a mixture of purified α 2M and rIL-6 generated three peaks corresponding to the α 2M-IL-6 complex, polymer of IL-6 and free IL-6. In contrast, a mixture of serum from α 2M-deficient PCa patients and rIL-6 did not demonstrate the peaks representing the α 2M-IL-6 complex and free IL-6. These results suggest that a function of α 2M is to bind and stabilize IL-6 by forming a complex. Based on these findings, IL-6 may be unstable in PCa patients with α 2M deficiency, leading to decreased production of CRP and SAA in liver cells. This is supported by the evidence that the serum levels of α 2M-dependent acute inflammatory biomarkers such as CRP, SAA and IL-6 were indeed decreased in PCa patients with α 2M deficiency.

Some other acute inflammatory biomarkers including α 1AT, α 1AG and CP are also known to increase in various conditions. It has been reported that α 1AT may be a significant factor in the differential diagnosis of serous effusion in patients with malignant disease (24). Yildirim *et al* reported that α 1AG is a useful prognostic factor in addition to performance status in patients with primary lung cancer (25). It has been also shown that CP may be a reliable marker for prostate cancer where is not accompanied by elevation of serum PSA (26). Further, Kasprzyk *et al* have reported that α 1AT, α 1AG and CP are useful prognostic factors in the risk assessment of non-small lung cancer recurrence after surgical management (27). In the present study, serum levels of α 1AT, α 1AG and CP in PCa patients with and without α 2M deficiency were significantly higher than in healthy controls. Moreover, there was no significant difference in levels

of these three factors between PCa patients with and without α 2M deficiency. These results suggest that α 1AT, α 1AG and CP are acute inflammatory biomarkers which are independent of α 2M function as a carrier protein of IL-6.

In conclusion, the present study demonstrated that α 2M can stabilize IL-6 by forming a complex, and in a mixture of serum from α 2M-deficient PCa patients and rIL-6, the α 2M-IL-6 complex and free IL-6 could not be detected. Furthermore, serum levels of CRP, SAA and IL-6, which are dependent on α 2M function, were not increased in PCa patients with α 2M deficiency. Therefore, measurement of the acute inflammatory biomarkers (CRP, SAA and IL-6) that depend on α 2M in combination with those independent of α 2M function (α 1AT, α 1AG and CP) in PCa patients may be an auxiliary indicator, together with PSA, to monitor PCa disease progression.

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